

Molecular Hydrogen: New Antioxidant and Anti-inflammatory Therapy for Rheumatoid Arthritis and Related Diseases

Toru Ishibashi*

Haradoi Hospital, Department of Rheumatology and Orthopaedic Surgery, 6-40-8 Aoba, Higashi-ku, Fukuoka 813-8588, Japan

Abstract: Rheumatoid arthritis (RA) is a chronic inflammatory disease in which the progressive destruction of joint causes morbidity. It is also associated with an increased risk of atherosclerosis, which can result in cardiovascular disease and mortality. The therapeutic goal is to control the systemic inflammation to obtain not only the remission of symptoms, but also improve general state of health. Although recent biologic immunosuppressive therapies targeting pro-inflammatory cytokines have spawned a paradigm shift regarding the prognosis of RA, these therapies possess inherent side effects. Also, early diagnosis of the disease remains confounded by uncertainty. While the mechanisms responsible for the onset of RA remain unclear, reactive oxygen species (ROS) play a significant role in the pathogenesis of RA. ROS play a central role both upstream and downstream of NF- κ B and TNF α pathways, which are located at the center of the inflammatory response. Among the ROS, the hydroxyl radical is the most harmful, and molecular hydrogen (H₂) is a selective scavenger for this species. Recently, it has been shown that H₂ is useful when administered along with the conventional therapy in RA as it acts to reduce oxidative stress in the patients. Especially in the early stage, H₂ showed significant therapeutic potential, which also seemed to assist diagnosis and treatment decisions of RA. The possible expectations regarding the potential benefits of H₂ by reducing the oxidative stress, resulting from inflammatory factors, are raised and discussed here. They include prevention of RA and related atherosclerosis, as well as therapeutic validity for RA.

Keywords: Rheumatoid, atherosclerosis, prevention, Oxidative Stress, 5 ppm, Molecular Hydrogen, 8-hydroxyguanine, Hydroxyl Radical.

INTRODUCTION

Rheumatoid arthritis (RA) is a chronic inflammatory disease that affects approximately 1% of the population. It is characterized by irreversible joint disorder accompanied by destruction of bone and cartilage, which causes serious morbidity. In addition, the chronic inflammation associated with RA can increase one's risk of atherosclerosis, which is a significant cause of mortality with the cardiovascular failure [1, 2]. Atherosclerosis associated with RA progresses rapidly, even in the absence of the conventional risk factors, such as hypertension, diabetes mellitus, or obesity. Consequently, the aim of RA therapy includes not only improving the disease activity, which is commonly estimated by the joint disorder and inflammatory markers, but also controlling the systemic and unremarkable inflammation of endothelial cells. Recent progress of anti-cytokine therapies is improving the risk for cardiovascular disease (CVD) [3].

Although the etiology is unknown, RA is certainly associated with autoimmune disorders, and its pathogenesis has been well investigated [2]. Auto-reactive T cells that infiltrate the synovial tissue promote the immune response, resulting in an overproduction of pro-inflammatory cytokines such as tissue necrosis factor alpha (TNF α), interleukin 1 (IL-1) and interleukin 6 (IL-6). Accordingly, early therapy was based on aggressive biological modification of the disease by controlling the synovial T cells and/or reducing the levels of the cytokines. Unfortunately, this approach has met limited therapeutic success, raising the issue that important regulatory factors were missing in the existing mechanistic model of RA. Reactive oxygen species (ROS) could be one of the unidentified regulatory factors. The synovial fluid and peripheral blood of RA patients have high levels of ROS and ROS-generated molecules, including superoxide, peroxide, hydroxyl radicals and reactive nitrogen species like peroxynitrite [4-6]. They oxidize various cellular and extracellular components, including nucleotides, DNA,

proteins, polysaccharides, and lipids, by means of their unpaired free radicals. Of these, 8-hydroxyguanine (8-OHdG), which is produced by the oxidation of guanine bases in DNA and also in the nucleotide pools, is considered important [7-9]. 8-OHdG is a standard biomarker for oxidative stress. Numerous studies have reported that 8-OHdG accumulates in diseases related to oxidative stress, such as cancer, diabetes mellitus, Alzheimer's disease, hypertension, cardiovascular disease, metabolic syndrome, and autoimmune disease [10-16]. Elevated levels of 8-OHdG have been reported in RA [17, 18] and atherosclerosis [19].

In the past decade, it has been shown that molecular hydrogen (H₂) selectively eliminates highly reactive hydroxyl radical in cultured cells and living organisms [20, 21]. H₂ targets hydroxyl radicals, but not super oxide, peroxide, or nitric oxide, which are important molecules for organisms [22]. Recently, it was demonstrated that consumption of water with a high concentration of molecular hydrogen (4-5 ppm in the water) significantly improves the disease activity and reduces the oxidative stress in RA [23]. H₂ seemed to complement or provide a substitute for conventional therapy by reducing oxidative stress and improving damage associated with RA, especially in the early stages of the disease and in the case of Antibodies against cyclic citrullinated peptide (ACPA)-negative RA.

In this review article, prospective applications of new H₂ therapies, for both the diagnosis and treatment of RA, are discussed. Also the possible expectation for the prevention of RA and related atherosclerosis by the daily consumption of high H₂ water are mentioned.

GENERATION OF ROS IN CHRONIC INFLAMMATION

ROS are produced as an inevitable byproduct of electron transfer in oxidative phosphorylation during aerobic metabolism [24]. On the other hand, during inflammatory stages of RA, infiltration or proliferation of immune activated cells in the synovium actively generate ROS via the NADPH (nicotinamide adenine dinucleotide phosphate) oxidase system (Nox) [25-27]. Among the actively generated ROS, superoxide anion is the primary product and liberated into extracellular matrix as well as sequestered in lysosomes. Su-

*Address correspondence to this author at the Haradoi Hospital, Department of Rheumatology and Orthopaedic Surgery, 6-40-8 Aoba, Higashi-ku, Fukuoka 813-8588, Japan; Tel: +81-92-691-3881; Fax: +81-92-691-1059; E-mail: toruishi@haradoi-hospital.com

peroxide is then converted to hydrogen peroxide either spontaneously or catalytically by the superoxide dismutase (SOD) [28]. Hydrogen peroxide can then be converted to water by catalase (CAT) [29]. In the presence of iron (Fe^{2+}) or other transition metal ions, hydrogen peroxide is converted to hydroxyl radicals via the Fenton reactions [30]. Ferrous ions also have the ability to convert superoxide and hydrogen peroxide to hydroxyl radical by the Haber-Weiss reaction [31]. Reduction system of ROS is also carried by another mechanism that involves GPx and GST; this mechanism requires reduced glutathione (GSH) as a cofactor. Glutathione peroxidase (GPx) reduce hydrogen peroxide to water [29] and glutathione S-transferase (GST) catalyzes various detoxifications by conjugating GSH, including ROS [32]. Altogether, these processes for ROS generation, which start from superoxide production by Nox system, play an important role in defense mechanisms. At the same time, these defense mechanisms are equipped with SOD, CAT, GPx, and GST, to compensate for the toxicity resulting from ROS. ROS produced by severe or chronic inflammation may exceed the capability of the corresponding anti-oxidant enzymes, which seems to cause imbalance in the redox state [33]. Especially under such conditions, hydroxyl radical is unrestrictedly surplus because specific detoxification system is not known to exist in living cells. It is likely to be responsible for the majority of cytotoxicity associated with ROS due to its rapid and indiscriminate reactivity [34].

ROLE OF ROS IN REDOX REGULATION OF NF- κ B-DEPENDENT INFLAMMATORY CASCADES IN RA

Oxidative stress regulates several cellular processes, including the signaling pathways responsible for inflammatory responses. The activation status of signaling intermediates is regulated by ROS/GSH-mediated modifications of their thiol (R-SH) groups [33]. Excess ROS production disrupts this redox balance and amplifies inflammatory responses via NF- κ B. This leading transcription factor induces the transcription of several pro-inflammatory cytokines. All of the ROS described above cause an imbalance of redox state within the inflamed tissue, resulting in the activation of NF- κ B and related transcription factors. NF- κ B is a central regulator of cellular inflammatory response as it controls many of the genes involved in inflammation [35, 36]. It induces the transcription of several pro-inflammatory cytokines including TNF α , IL-1, and IL-6, which play key roles in the progression of RA and therefore are therapeutic drug targets. Among these, TNF α plays a pivotal role in the persistent inflammation of RA synovium, through its stimulation of oxidative stress [36, 37]. TNF α , produced downstream of the NF- κ B transcriptional complex, re-activates NF- κ B by releasing it from I- κ B, which in steady state phosphorylates and inactivates NF- κ B by forming complex with this transcription factor [38, 39]. In this manner, the "Loop 1" acts as a positive feedback mechanism within the NF- κ B and TNF α system and promotes the inflammatory response (see the "Loop 1" in Fig. 1).

In the synovial tissue, high levels of the pro-inflammatory cytokines activate and recruit neutrophils, synovial fibroblasts, and macrophages, which contribute to further overproduction of ROS by the Nox system. This ROS re-enters the pathway, upstream of NF- κ B, creating another positive feedback loop within the synovial inflammation pathway (see the "Loop 2" in Fig. 1). These loops stimulate the production of ROS and pro-inflammatory cytokines, both of which contribute to the accumulation of oxidative stress.

In addition to the oxidative stress, the NF- κ B also stimulates proteolytic activity. Proteolytic enzymes play the central role in the destruction of cartilage and bone associated with RA. Cytokines including TNF α or IL-1 activate synovial fibroblasts and macrophages to produce cartilage-destroying enzymes, such as metalloproteinases (MMP), including MMP-1 and MMP-3. They are thought to be secreted from the synovial fibroblasts and macrophages especially in the pannus of advanced rheumatoid joints and

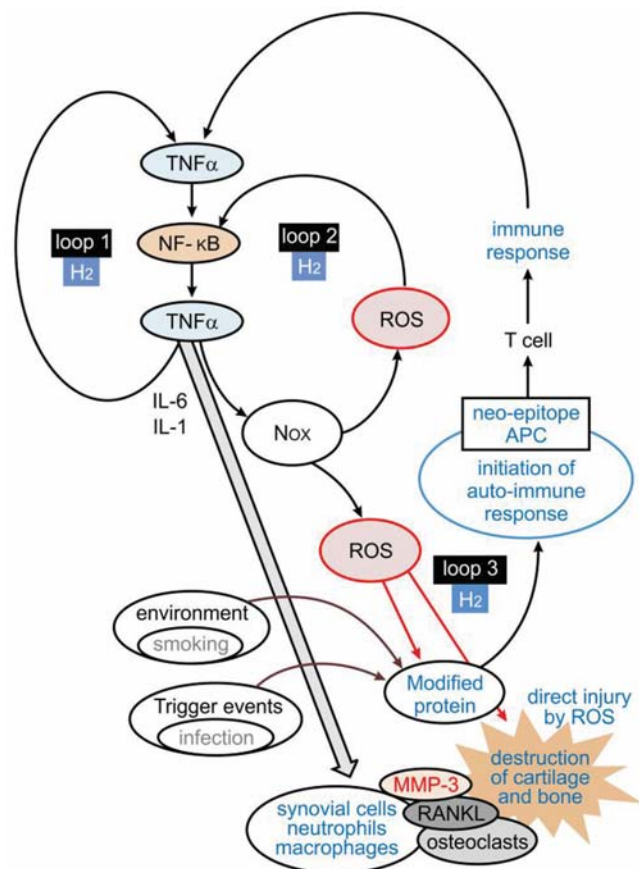


Fig. (1). A schematic representation of the three loops involved in amplification of inflammation in patients with RA and related atherosclerosis. Loop 1 indicates the NF- κ B-TNF α positive feedback loop. Loop 2 indicates the redox sensing loop by ROS-NF- κ B-TNF α . Both loops can be blocked by using H₂ that scavenges hydroxyl radicals directly or via NF- κ B pathways. ROS, which are generated by Nox system and amplified through these loops, then stimulate synovial fibroblasts, neutrophils, and macrophages, which promote cartilage and bone erosion via MMP-3 or RANKL expression. In addition, the modified proteins by ROS may generate a loop 3 which may promote the autoimmunity response by feeding back into loops 1 and 2.

can degrade hyaluronic acid, proteoglycan, and collagen which construct cartilage [40]. Also, chondrocytes stimulated by IL-1 secrete MMP-3 and promote the absorption of the subchondral bones [41, 42]. MMP-3 is considered an important protease in RA joint destruction as the concentration of MMP-3 in the serum of RA patients is correlated with disease activity [43]. In addition, the activation of TNF α leads to the elevated expression of molecules, such as RANKL (receptor activator of NF- κ B ligand), related to bone-destruction [44]. Expression of RANKL in the synovial fibroblasts results in differentiation of the osteoclast progenitor cells expressing RANK on their surface. The progenitor cells, including monocytes, which infiltrate in the synovial tissue, mature into osteoclasts—the primary cells responsible for the absorption of bone—which are responsible for the bone erosion associated with RA [2, 45]. Again, ROS located upstream of these cytokines, are indirectly responsible for the destruction of cartilage and bone in the joints of RA patients (Fig. 1). It has also been discussed that the synovial fluid in RA is hypoxic and the synovial reperfusion seems to stimulate the redox cascade including NF- κ B [36].

Recent successful anti-inflammatory therapies targeting TNF α seem to be attributed to disrupting the inflammatory Loop 1. These findings suggest that a therapeutic approach targeting the ROS-TNF α amplification loop would significantly reduce inflammation-mediated damage in the joints of RA patients. As evident in (Fig. 1), such a therapy that breaks down the ROS feedback loop may be effective and necessary for preventing damage associated with RA.

DIRECT DAMAGE OF JOINT TISSUE BY ROS

Proteolytic activity is not the only source of cartilage and bone destruction associated with RA. ROS also have direct effects as they oxidize and degrade the major components of cartilage and bone, including collagen and hyaluronic acid (HA) [6]. Numerous studies have detected products of ROS reactivity in serum as well as in the joint fluids from patients with RA [5]. The presence of hyaluronic acid in reduced state and abundant leukocyte infiltration are common features of RA joint fluid. The degradation of HA is, at least in part, the result of fragmentation by hydroxyl radical [6, 46, 47]. The decreased viscosity of joint fluid in RA patients is thought to be attributed to such degradation of HA. Hydroxyl radicals also modify collagen, resulting in the formation of cross-links or fragmentation of the protein [48, 49]. Such degraded collagen seems to be more susceptible to subsequent cleavage by proteinase in the joint fluid.

These degrading processes and subsequent tissue remodeling of the synovial tissue could invite phagocytic and proteinase activities, which result in the induction of ROS-cytokine activated inflammatory 'Loop 1 and Loop 2', as shown in (Fig. 1). The positive feedback resulting from the accumulation of oxidized products could further exacerbate the cartilage and bone erosion, and lead to joint destruction. Therefore, protection of the joint components from the direct degradation by ROS represents another potential target of RA therapy.

GENETIC ASSOCIATION WITH RA AND ROLE OF ROS IN THE ETIOLOGY

Genetic modifications in the human leukocyte antigen (HLA) region account for nearly half of the genetically-related risk factors of RA. For instance, patients carrying the shared epitope (SE)—the region encoding QKRAA, QRRAA, or RRRRAA amino acid sequences—in the third hyper-variable region of the human leukocyte antigen-DRB1 (HLA-DRB1) have a high susceptibility to develop RA [50, 51]. The risk for RA was reported as 15% for monozygotic twins carrying the SE [52], compared to 1% in the general population. This motif predicts not only the susceptibility to RA but also the severity of prognosis [51]. Recently it has been postulated that the combination of SE with other factors such as genetic polymorphisms and/or environment interact to increase the susceptibility to RA. First, it has been reported that significant positive interactions between HLA-DRB1-SE and smoking and the auto-antibody named ACPA incur the highest risk for the onset of RA [53, 54]. Second, functional mutations in the antioxidant enzyme GST are involved in increasing the genetic susceptibility of RA patients as they increase oxidative stress. The GST Mu-1 (GSTM1-null) genotype, which results in complete inhibition of the enzyme activity, exerts significant additive interactions with HLA-DRB1-SE on the risk of being ACPA-positive in RA patients [55]; this study provides evidence that oxidative stress increases susceptibility for the development of RA.

Oxidative stress may participate in the etiology of RA through direct interaction with DNA. For example, 8-OHdG is produced by the oxidation of guanine bases of DNA and is also present in the nucleotide pools [7-9]. It is highly mutagenic because it pairs with adenine as well as cytosine. This property causes transversion mutations during DNA replication. Consequently, the possibility exists that ROS may induce somatic mutations that result in altered protein function and/or immunogenicity and function as neo-epitopes

[56-58]. These could continue to activate the immune system [58, 59]. Elevated levels of 8-OHdG have been reported in RA [17, 18]. However, somatic mutations of genomic DNA do not appear to be responsible for the auto-antigen in RA. If they were, one would expect homogeneous and stable epitopes in the pocket of the APC in the synovial tissue; however, these have never been detected even after extensive research.

Another possibility is that ROS primes the immune response through effects on mitochondrial DNA (mtDNA). A somatic cell has multiple copies of mitochondria, which have individual genome susceptible to mutations caused by ROS [60-62]. Somatic mutations of mtDNA have been reported to be associated with RA [63]. There is a slightly greater chance that the mutations of mtDNA are responsible for priming the immune response because it would be more difficult to demonstrate. Possible mutations in mtDNA and five mutated peptides that could potentially represent a SE have been identified in RA patients [64].

Regarding the potential role of ROS in RA etiology, more common considerations involve processes downstream from the DNA. For example, the mutagenic nucleic acid, 8-hydroxyguanosine, also causes partial phenotypic suppression during transcription [65]. It is likely that the error-proteins would be identified as foreign molecules and function as neo-epitopes [5, 58]. The error-proteins with alternative post-translational modifications are also included in this hypothesis. Antibodies for the post-translationally modified proteins containing citrulline are highly detected, especially among RA patients with poor prognosis. Although the relationship between ROS and loss of tolerance to citrullinated protein remains unclear, structural alterations of peptide motifs caused by oxidative modification may play a role. Because those antigens are oxidized post-transcriptionally or post-translationally, they seem to be quite heterogeneous. If there is a possibility that such heterogeneous antigens could activate the immune response in a consistent manner, it seems that they must have a shared structure or electrochemically mutual characters, especially under the oxidative conditions. This hypothesis would establish a third loop, involving ROS, in the inflammation pathway (see Loop 3 in Fig. 1), involving priming of the auto-immune response to establish memory T cell clones against such neo-epitopes presented on antigen presenting cells (APC). In the state, down regulation of ROS seems to decrease the neo-epitopes of RA.

ATHEROSCLEROSIS ASSOCIATED WITH RA

Atherosclerosis is a common co-morbidity of RA, and a major cause of cardiovascular diseases (CVDs) and increased mortality among RA patients [66, 67]. While the pathogenesis of atherosclerosis usually involves the classical risk factors, including hypertension, obesity, or smoking, atherosclerosis associated with RA often occurs independent of these factors. Instead, the inflammatory cascades common to both RA and atherosclerosis, including pro-inflammatory cytokines and ROS appear responsible for the association between these disorders (see schematic Fig. 2). These inflammatory pathways affect the endothelial structure in blood vessels as well as synovial tissues in RA. Superoxide, produced by endothelial cells and smooth muscle cells via the Nox pathways including Nox1, Nox2, Nox4, and Nox5, are thought to be involved in endothelial dysfunction and the ROS-related progress of atherosclerosis [68-71]. In the case of atherosclerosis, oxidized LDL, located downstream of the intersection of these pathways, induces the formation of plaque responsible for the increased risk of CVDs [72, 73]. The development of atherosclerosis in RA patients is initiated by endothelial phenotypic alterations in response to large amounts of noxious stimuli. The increased expression of adhesion molecules such as intercellular adhesion molecules 1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and E-selectin, the enhancement of pro-inflammatory cytokines (TNF- α , IL-1, interferon- γ), and the increase in oxidative stress initiate this condition

[33]. It was reported that the up-regulation of TNF α expression, alone, can cause vascular dysfunction [74]. In healthy volunteers, intra-arterial administration of TNF α , at a dose of 80 or 240 ng/min for 30 min, resulted in an acute vascular inflammation associated with impaired endothelial structure. Recently, it has been reported that anti-TNF α therapies could improve the progression of atherosclerosis in RA patients [3, 75]. These results indicate that the pathogenesis of atherosclerosis in RA is caused by the shared TNF α /ROS inflammatory pathway at the crossing between Loop 1 and 2 as shown in (Fig. 2). Accordingly, ROS should represent a suitable therapeutic target for the treatment of both RA and associated atherosclerosis.

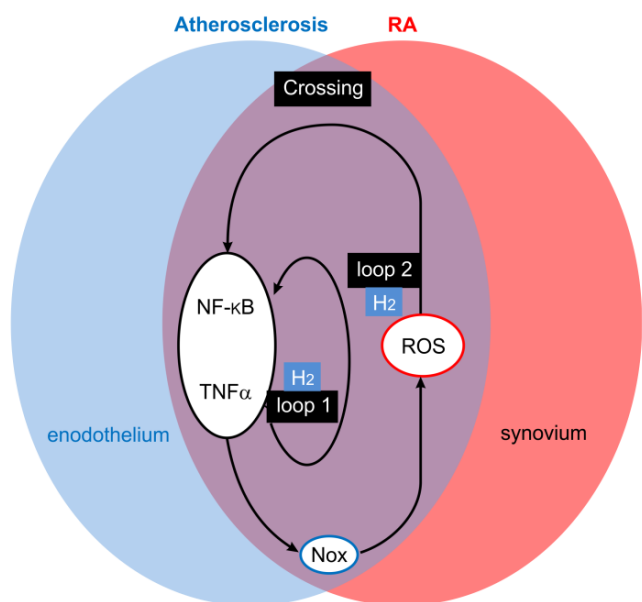


Fig. (2). A schematic representation of the inflammatory cascades common to both RA and atherosclerosis. This common inflammatory cascade includes both loop 1 and loop 2, thereby amplifying the positive feedback reactions. Atherosclerosis is believed to be complicated by inflammation resulting from RA.

THERAPEUTIC POTENTIAL OF MOLECULAR HYDROGEN (H₂) FOR THE TREATMENT OF RA

The present review highlighted the multiple roles of ROS in the development of RA and atherosclerosis, as toxic damaging agents, in the amplification of NF- κ B-dependent inflammatory response, and as mutagenic agents. Accordingly, new therapeutic approaches should target the reduction of ROS in circulation and in the joints of RA patients. In the past 30 years, several clinical studies evaluated the potential of antioxidant therapies for RA using predominantly synthetic forms of SOD or the ROS scavenger edaravone [76-79]. However, these results do not clearly establish whether these scavengers effectively reduce the disease activity in patients with RA. Further, none of them targeted hydroxyl radical, which seems to be responsible for most of the damage caused by ROS. Unlike superoxide, which can be eliminated by SOD, the hydroxyl radical cannot be eliminated by an enzyme. Consequently, the antioxidant strategies envisioned so far were not addressing the most destructive member of the ROS family. Considering the central involvement of ROS in the inflammatory pathways of RA and that the hydroxyl radical is the most toxic species, one would expect therapies that target this radical for effective treatment.

One antioxidant that has shown promising results is molecular hydrogen (H₂). Molecular hydrogen reacts with and detoxifies the hydroxyl radical without forming other radicals [21, 80]. Within the past decade, H₂ has been explored in the treatment of oxidative

stress. Numerous studies in animals have shown the therapeutic potential of H₂ for diseases associated with ROS [22], and has already been used in some clinical treatment, such as type II diabetes mellitus, metabolic syndrome, hemodialysis, muscular diseases, and acute brain stem infarction [81-85]. The therapeutic potential of H₂ was first shown using the model for schistosomiasis-associated chronic liver inflammation. In the study, livers of schistosomiasis-infected mice were protected from the chronic inflammation by the treatment with 0.7 MPa hydrogen for two weeks [21]. Furthermore, it has also been shown that H₂ selectively eliminates the hydroxyl radical in cultured cells and living organisms [20]. In addition to its therapeutic potential for the elimination of hydroxyl radicals, H₂ is also used by deep-sea divers to prevent the decompression sickness, where its safety has been established [86, 87]. During these practical and experimental applications of H₂, no adverse effects have been reported.

Recently we reported that H₂ is the effective molecule not only for a compensational use along with insufficient anti-rheumatic drugs but also for the diagnostic or monotherapeutic use at early stage in patients with RA [23]. In the study, patients were administered water containing 4-5 ppm H₂ (high H₂ water). By drinking 500 ml of the high H₂ water daily for four weeks, oxidative stress was effectively reduced and the disease activity in RA was significantly improved. H₂ seemed to have complemented conventional RA therapy by reducing oxidative stress. At least in the early stage of disease progression and among ACPA-negative patients, H₂ seems to have a potential to assist with diagnosis and treatment of non-aggressive or transient RA. Also considering the crossing mechanisms shown in (Fig. 2), we expect that the development of atherosclerosis in RA patients could be slowed or even prevented by the daily uptake of H₂ (shown in Fig. 2).

Many questions regarding the effectiveness of H₂ for the treatment of RA still remain. Among the concerns expressed by experts in the field are the short residence time and the extremely low amount of H₂ administered by drinking water with 1.6 ppm H₂ at the highest than the inhaled 2% H₂ gas. This discrepancy of more than 100 times of difference between them, even if they seem to have similar biological effects on organisms, have been discussed previously [80]. The authors proposed the possibility of H₂ for a gaseous signal modulator to explain the unknown mechanisms for H₂ effects. Another concern is related to alterations of gene expression that have been measured in response to H₂, using DNA microarray analysis [88]. In this study, H₂ caused the up-regulation of 548 and down-regulation of 695 genes in rat liver. Among these, the up-regulation of redox-related genes was especially prominent. The results raised the possibility that, while minimal in magnitude, numerous effects on the gene expression caused by H₂ may shift indirect molecular reactions in an organism. Furthermore, additional mechanisms related to the products of hydroxyl radical are still unknown. I discuss some of them in the next section.

EXPECTATIONS REGARDING THE USE OF H₂ FOR THE PREVENTION OF RA AND ATHEROSCLEROSIS

In our study, H₂ seems to have an influence even during the washout period when the patients did not drink high H₂ water for 4 weeks [23]. The persistence of this effect cannot be explained by continuous elimination of hydroxyl radical. It is believed that H₂ reach into cytosol, nucleus and even into mitochondria of various tissues by gaseous diffusion [22]. H₂ administered via high H₂ water as well as H₂-saturated water (containing 1.6ppm H₂) rapidly reached the maximum concentration in the lung within 10 min and was exhaled within 60 min [23]. This indicates that the surplus H₂ passed through the body within a small period, except for the H₂, which seems to have reacted with hydroxyl radical. The removal of hydroxyl radicals and interference with the inflammatory response pathways described above likely resulted from exposure to H₂; however, other explanations are required to explain the continuation

of benefits observed once H₂ administration had been terminated. The effects by hydroxyl radicals are rapid and detrimental, but the target molecules oxidized by hydroxyl radicals may carry the 'reactive' signal until it is degraded. Constitutive stimulation by such oxidized molecules may be eligible for the chronic inflammation of RA. One of such molecules is 8-OHdG, but other molecules that have been unknown or neglected should be investigated in the future.

By using H₂ therapeutically for RA, some questions regarding the etiology of RA may be resolved. If modified antigen is produced by hydroxyl radical and the immunogenic molecules with the shared epitope are responsible for the onset of RA, H₂ could eliminate the immunogenic state during the early stages of RA, resulting in only transient inflammation. In such cases, constitutive consumption of H₂ would not be required, at least until the next trigger events such as infections or environmental stress occur. Alternatively, if the constitutive consumption of H₂ would be required to improve RA even at the early stage, H₂ seems to be effective except for the 'Loop 3' in (Fig. 1). Both of these schemas are possible. As it is widely believed that RA is divided into subclasses based on its mechanisms of onset or the prognosis, the efficacy of H₂ in the treatment of RA may differ among the subclasses of RA.

Altogether, it is conceivable that H₂ therapy has individual window of opportunity. The therapeutic window of opportunity is critical in determining the prognosis of RA, as usually observed with the conventional immunosuppressive drugs. If this window is missed, inflammation can advance and cause joint and bone destruction. The window for H₂ therapy will depend on the stages where H₂ works to reduce the inflammation, as shown in (Fig. 1). In cases where H₂ therapy misses the opportunities, it may have only restricted effect on the inflammation of RA.

The recent success of and the paradigm shift within drug therapies seem to ignore the efforts to understand the origin of RA. When the precise relationship between RA and ROS is observed by H₂, we have to remember the final goal to exterminate the auto-antigen responsible for RA.

Further study is required to confirm the therapeutic effectiveness of H₂ on RA. Randomized, double blinded, and placebo-controlled investigation of the effects of high H₂ water and the infusion of H₂-resolved saline are ongoing (data not shown). We expect that H₂ therapy in very early stage could prevent the onset of RA, if it is able to interfere with Loop 3 at the onset of the auto-immune response by reducing the ROS-related neo-epitope. Alternatively, the eventual activation of pro-inflammatory cytokines, which may trigger the auto-immune response, may be prevented by daily H₂ consumption.

CONCLUSION

H₂ is an inert gas present within the human body and is not classified as a medicine, but it has been shown to have therapeutic and diagnostic potential for RA as discussed here. The apparatus for creating H₂-enriched water (over 5 ppm of H₂) is already available commercially. The preventive effect of H₂ may be shown by those populations who drink the high H₂ water daily. Also, additional investigations of the benefits of H₂ for RA patients are urgently needed, as H₂ therapy has tremendous potential, but is currently under-utilized and its benefits are not thoroughly explored.

CONFLICT OF INTEREST

The author confirms that this article content has no conflicts of interest.

ACKNOWLEDGEMENTS

I thank Bunpei Sato, Mami Nagao, Yuichi Hara, Yuji Naritomi, Hiroshi Hara, and Tetsuhiko Nagao for their support during the

preparation of the manuscript. I am also grateful to Kota Sasaki and Kazuhisa Fukuoka for their excellent advice.

ABBREVIATIONS

8-OHdG	=	8-hydroxydeoxyguanine
ACPA	=	Antibodies against cyclic citrullinated peptide
DAS28	=	Disease activity score in 28 joints
APC	=	Antigen presenting cells
CVD	=	Cardiovascular disease
Nox	=	Nicotinamide adenine dinucleotide phosphatase
CAT	=	Catalase
GSH	=	Reduced glutathione
GPx	=	Glutathione peroxidase
GST	=	Glutathione S-transferase
HLA	=	Human leukocyte antigen
ROS	=	Reactive oxygen species
RA	=	Rheumatoid arthritis
SOD	=	Superoxide dismutase
TNF α	=	Tumor necrosis factor α
NF- κ B	=	Nuclear factor kappa B
MtDNA	=	Mitochondrial DNA
RANK	=	Receptor activator of NF- κ B
ICAM-1	=	Intercellular adhesion molecules-1
VCAM-1	=	Vascular cell adhesion molecule-1

REFERENCES

- [1] Smith HS, Smith AR, Seidner P. Painful rheumatoid arthritis. *Pain Physician*. 2011; 14: E427-58.
- [2] McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med* 2011; 365: 2205-19.
- [3] Gonzalez-Juanatey C, Vazquez-Rodriguez TR, Miranda-Filloo JA, *et al.* Anti-TNF-alpha-adalimumab therapy is associated with persistent improvement of endothelial function without progression of carotid intima-media wall thickness in patients with rheumatoid arthritis refractory to conventional therapy. *Mediators Inflamm* 2012 doi: 10.1155/2012/674265
- [4] Filippin LI, Verdelino R, Marroni NP, Xavier RM. Redox signaling and the inflammatory response in rheumatoid arthritis. *Clin Exp Immunol* 2012; 152: 415-22.
- [5] Winyard PG, Ryan B, Eggleton P, *et al.* Measurement and meaning of markers of reactive species of oxygen, nitrogen and sulfur in healthy human subjects and patients with inflammatory joint disease. *Biochem Soc Trans* 2011; 39: 1226-32.
- [6] Hadjigogos K. The role of free radicals in the pathogenesis of rheumatoid arthritis. *Panminerva Med* 2003; 45: 7-13.
- [7] Kasai H, Crain PF, Kuchino Y, Nishimura S, Ootsuyama A, Tanooka H. Formation of 8-hydroxyguanine moiety in cellular DNA by agents producing oxygen radicals and evidence for its repair. *Carcinogenesis* 1986; 7: 1849-51.
- [8] Maki H, Sekiguchi M. MutT protein specifically hydrolyses a potent mutagenic substrate for DNA synthesis. *Nature* 1992; 355: 273-5.
- [9] Ishibashi T, Hayakawa H, Sekiguchi M. A novel mechanism for preventing mutations caused by oxidation of guanine nucleotides. *EMBO Rep* 2003; 4: 479-83.
- [10] Negishi H, Ikeda K, Kuga S, *et al.* The relation of oxidative DNA damage to hypertension and other cardiovascular risk factors in Tanzania. *J Hypertens* 2001; 19: 529-33.
- [11] Nishikawa T, Sasahara T, Kiritoshi S, *et al.* Evaluation of urinary 8-hydroxydeoxy-guanosine as a novel biomarker of macrovascular complications in type 2 diabetes. *Diabetes Care* 2003; 26: 1507-12.
- [12] Shen J, Gammon MD, Terry MB, *et al.* Telomere length, oxidative damage, antioxidants and breast cancer risk. *Int J Cancer* 2009; 124: 1637-43.

- [13] Bashir S, Harris G, Denman MA, Blake DR, Winyard PG. Oxidative DNA damage and cellular sensitivity to oxidative stress in human autoimmune diseases. *Ann Rheum Dis* 1993; 52: 659-66.
- [14] Miyamoto M, Kotani K, Ishibashi S, Taniguchi N. The relationship between urinary 8-hydroxydeoxyguanosine and metabolic risk factors in asymptomatic subjects. *Medical Principles Pract* 2001; 20: 187-90.
- [15] Masugata H, Senda S, Murao K, *et al.* Association between urinary 8-Hydroxydeoxyguanosine, an indicator of oxidative stress, and the cardio-ankle vascular index in hypertensive patients. *J Atheroscler Thromb* 2012; 19: 747-55.
- [16] Mao P, Manczak M, Calkins MJ, *et al.* Mitochondria-targeted catalase reduces abnormal APP processing, amyloid beta production and BACE1 in a mouse model of Alzheimer's disease: implications for neuroprotection and lifespan extension. *Hum Mol Gen* 2012; 21: 2973-90.
- [17] Kageyama Y, Takahashi M, Ichikawa T, Torikai E, Nagano A. Reduction of oxidative stress, and the cardio-ankle vascular index in hypertensive marker levels by anti-TNF-alpha antibody, infliximab, in patients with rheumatoid arthritis. *Clin Exp Rheumatol* 2008; 26: 73-80.
- [18] Rall LC, Roubenoff R, Meydani SN, Han SN, Meydani M. Urinary 8-hydroxy-2'-deoxyguanosine (8-OHdG) as a marker of oxidative stress in rheumatoid arthritis and aging: effect of progressive resistance training. *J Nutr Biochem* 2000; 11: 581-4.
- [19] Wu LL, Chiou CC, Chang PY, Wu JT. Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics. *Clin Chim Acta* 2004; 339: 1-9.
- [20] Ohsawa I, Ishikawa M, Takahashi K, *et al.* Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. *Nature Med* 2007; 13: 688-94.
- [21] Gharib B, Hanna S, Abdallahi OM, Lepidi H, Gardette B, De Reggi M. Anti-inflammatory properties of molecular hydrogen: investigation on parasite-induced liver inflammation. *C R Acad Sci III* 2001; 324: 719-24.
- [22] Ohta S. Recent progress toward hydrogen medicine: potential of molecular hydrogen for preventive and therapeutic applications. *Curr Pharm Des* 2011; 17: 2241-52.
- [23] Ishibashi T, Sato B, Rikitake M, *et al.* Consumption of water containing a high concentration of molecular hydrogen reduces oxidative stress and disease activity in patients with rheumatoid arthritis: an open-label pilot study. *Med Gas Res* 2012; 2: 27.
- [24] Takahashi M. Oxidative stress and redox regulation on *in vitro* development of mammalian embryos. *J Reprod Dev* 2012; 58: 1-9.
- [25] Chenevier-Gobeaux C, Lemarchal H, Bonnefont-Rousselot D, Poiraudou S, Ekindjian OG, Borderie D. Superoxide production and NADPH oxidase expression in human rheumatoid synovial cells: regulation by interleukin-1 beta and tumour necrosis factor-alpha. *Inflamm Res* 2006; 55: 483-90.
- [26] Griffiths HR. Is the generation of neo-antigenic determinants by free radicals central to the development of autoimmune rheumatoid disease? *Autoimmun Rev* 2008; 7: 544-9.
- [27] Kundu S, Ghosh P, Datta S, Ghosh A, Chattopadhyay S, Chatterjee M. Oxidative stress as a potential biomarker for determining disease activity in patients with Rheumatoid Arthritis. *Free Radic Res* 2012; 46: 1482-9.
- [28] Turrens JF. Mitochondrial formation of reactive oxygen species. *J Physiol* 2003; 552: 335-44.
- [29] Droge W. Free radicals in the physiological control of cell function. *Physiol Rev* 2002; 82: 47-95.
- [30] Sheu SS, Nauduri D, Anders MW. Targeting antioxidants to mitochondria: a new therapeutic direction. *Biochim Biophys Acta* 2006; 762: 256-65.
- [31] Puppo A, Halliwell B. Formation of hydroxyl radicals in biological systems. Does myoglobin stimulate hydroxyl radical formation from hydrogen peroxide? *Free Radic Res Commun* 1988; 4: 415-22.
- [32] Yang Y, Parsons KK, Chi L, Malakauskas SM, Le TH. Glutathione S-transferase-micro1 regulates vascular smooth muscle cell proliferation, migration, and oxidative stress. *Hypertension* 2009; 54: 1360-8.
- [33] Kondo T, Hirose M, Kageyama K. Roles of oxidative stress and redox regulation in atherosclerosis. *J Atheroscler Thromb* 2009; 16: 532-8.
- [34] Candeias LP, Patel KB, Stratford MR, Wardman P. Free hydroxyl radicals are formed on reaction between the neutrophil-derived species superoxide anion and hypochlorous acid. *FEBS Lett* 1993; 333: 151-3.
- [35] Gulati P, Klohn PC, Krug H, Göttlicher M, Markova B, Böhmer FD, Herrlich P. Redox regulation in mammalian signal transduction. *IUBMB Life* 2001; 52: 25-8.
- [36] Hitchon CA, El-Gabalawy HS. Oxidation in rheumatoid arthritis. *Arthritis Res Ther* 2004; 6: 265-78.
- [37] Williams RO, Feldmann M, Maini RN. Anti-tumor necrosis factor ameliorates joint disease in murine collagen-induced arthritis. *Proc Nat Acad Sci USA* 1992; 89: 9784-8.
- [38] Lawrence T, Bebiec M, Liu GY, Nizet V, Karin M. IKK alpha limits macrophage NF-kappaB activation and contributes to the resolution of inflammation. *Nature* 2005; 434: 1138-43.
- [39] Lee EG, Boone DL, Chai S, *et al.* Failure to regulate TNF-induced NF-kappaB and cell death responses in A20-deficient mice. *Science* 2000; 289: 2350-4.
- [40] Klareskog L, Catrina AI, Paget S. Rheumatoid arthritis. *Lancet* 2009; 373: 659-72.
- [41] Schett G, Stach C, Zwerina J, Voll R, Manger B. How antirheumatic drugs protect joints from damage in rheumatoid arthritis. *Arthritis Rheum* 2006; 58: 2936-48.
- [42] Schett G. Review: Immune cells and mediators of inflammatory arthritis. *Autoimmunity* 2008; 41: 224-9.
- [43] Urata Y, Uesato R, Tanaka D, Nakamura Y, Motomura S. Treating to target matrix metalloproteinase 3 normalisation together with disease activity score below 2.6 yields better effects than each alone in rheumatoid arthritis patients: T-4 Study. *Ann Rheum Dis* 2012; 71: 534-40.
- [44] Lee SH, Kim TS, Choi Y, Lorenzo J. Osteoimmunology: cytokines and the skeletal system. *BMB Reports* 2008; 41: 495-510.
- [45] Hitchon CA, El-Gabalawy HS. The synovium in rheumatoid arthritis. *Open Rheumatol J* 2011; 5: 107-14.
- [46] Grootveld M, Henderson EB, Farrell A, Blake DR, Parkes HG, Haycock P. Oxidative damage to hyaluronate and glucose in synovial fluid during exercise of the inflamed rheumatoid joint. Detection of abnormal low-molecular-mass metabolites by proton-m.r. spectroscopy. *Biochem J* 1991; 273: 459-67.
- [47] Uchiyama H, Dobashi Y, Ohkouchi K, Nagasawa K. Chemical change involved in the oxidative reductive depolymerization of hyaluronic acid. *J Biol Chem* 1990; 265: 7753-9.
- [48] Hawkins CL, Davies MJ. Oxidative damage to collagen and related substrates by metal ion/hydrogen peroxide systems: random attack or site-specific damage? *Biochim Biophys Acta* 1997; 1360: 84-96.
- [49] Deberg M, Labasse A, Christgau S, *et al.* New serum biochemical markers (Coll 2-1 and Coll 2-1 NO2) for studying oxidative-related type II collagen network degradation in patients with osteoarthritis and rheumatoid arthritis. *Osteoarthritis Cartilage* 2005; 13: 258-65.
- [50] Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum* 1987; 30: 1205-13.
- [51] Holoshitz J. The rheumatoid arthritis HLA-DRB1 shared epitope. *Curr Opin Rheumatol* 2010; 22: 293-8.
- [52] Silman AJ, MacGregor AJ, Thomson W, *et al.* Twin concordance rates for rheumatoid arthritis: results from a nationwide study. *Brit J Rheumatol* 1993; 32: 903-7.
- [53] Klareskog L, Stolt P, Lundberg K, *et al.* A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheum* 2006; 54: 38-46.
- [54] Willemze A, van der Woude D, Ghiddey W, *et al.* The interaction between HLA shared epitope alleles and smoking and its contribution to autoimmunity against several citrullinated antigens. *Arthritis Rheum* 2011; 63: 1823-32.
- [55] Mikuls TR, Gould KA, Bynote KK, *et al.* Anti-citrullinated protein antibody (ACPA) in rheumatoid arthritis: influence of an interaction between HLA-DRB1 shared epitope and a deletion polymorphism in glutathione S-transferase in a cross-sectional study. *Arthritis Res Ther* 2010; 12: R213.
- [56] Phillips DC, Dias HK, Kitas GD, Griffiths HR. Aberrant reactive oxygen and nitrogen species generation in rheumatoid arthritis (RA): causes and consequences for immune function, cell survival, and therapeutic intervention. *Antioxid Redox Signal* 2010; 12: 743-85.

- [57] Tak PP, Zvaifler NJ, Green DR, Firestein GS. Rheumatoid arthritis and p53: how oxidative stress might alter the course of inflammatory diseases. *Immunol Today* 2000; 21: 78-82.
- [58] Bang H, Egerer K, Gauliard A, *et al.* Mutation and citrullination modifies vimentin to a novel autoantigen for rheumatoid arthritis. *Arthritis Rheum* 2007; 56: 2503-11.
- [59] Remans PH, van Oosterhout M, Smeets TJ, *et al.* Intracellular free radical production in synovial T lymphocytes from patients with rheumatoid arthritis. *Arthritis Rheum* 2005; 52: 2003-9.
- [60] Liu P, Demple B. DNA repair in mammalian mitochondria: Much more than we thought? *Environ Mol Mutagen* 2010; 51: 417-26.
- [61] Falkenberg M, Larsson NG, Gustafsson CM. DNA replication and transcription in mammalian mitochondria. *Annual Rev Biochem* 2007; 76: 679-99.
- [62] Schriener SE, Linford NJ, Martin GM, *et al.* Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* 2005; 308: 1909-11.
- [63] Da Sylva TR, Connor A, Mburu Y, Keystone E, Wu GE. Somatic mutations in the mitochondria of rheumatoid arthritis synoviocytes. *Arthritis Res Ther* 2005; 7: R844-51.
- [64] Ospelt C, Gay S. Somatic mutations in mitochondria: the chicken or the egg? *Arthritis Res Ther* 2005; 7: 179-80.
- [65] Ishibashi T, Hayakawa H, Ito R, Miyazawa M, Yamagata Y, Sekiguchi M. Mammalian enzymes for preventing transcriptional errors caused by oxidative damage. *Nucleic Acids Res* 2005; 33: 3779-84.
- [66] Cavagna L, Boffini N, Cagnotto G, Inverardi F, Grosso V, Caporali R. Atherosclerosis and rheumatoid arthritis: more than a simple association. *Mediators Inflamm* 2012; doi: 10.1155/2012/147354
- [67] Ozbalkan Z, Efe C, Cesur M, *et al.* An update on the relationships between rheumatoid arthritis and atherosclerosis. *Atherosclerosis* 2010; 212: 377-82.
- [68] Takac I, Schroder K, Brandes RP. The Nox family of NADPH oxidases: friend or foe of the vascular system? *Curr Hypertens Rep* 2012; 14: 70-8.
- [69] Montezano AC, Touyz RM. Reactive oxygen species and endothelial function--role of nitric oxide synthase uncoupling and Nox family nicotinamide adenine dinucleotide phosphate oxidases. *Basic Clin Pharmacol Toxicol* 2012; 110: 87-94.
- [70] Al Ghoulh I, Khoo NK, Knaus UG, *et al.* Oxidases and peroxidases in cardiovascular and lung disease: new concepts in reactive oxygen species signaling. *Free Radic Biol Med* 2011; 51: 1271-88.
- [71] Bedard K, Krause KH. The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiol Rev* 2007; 87: 245-313.
- [72] Carr AC, McCall MR, Frei B. Oxidation of LDL by myeloperoxidase and reactive nitrogen species: reaction pathways and antioxidant protection. *Arterioscler Thromb Vasc Biol* 2000; 20: 1716-23.
- [73] Yoshida H, Kisugi R. Mechanisms of LDL oxidation. *Clin Chim Acta* 2010; 411: 1875-82.
- [74] Chia S, Qadan M, Newton R, Ludlam CA, Fox KA, Newby DE. Intra-arterial tumor necrosis factor- α impairs endothelium-dependent vasodilatation and stimulates local tissue plasminogen activator release in humans. *Arterioscler Thromb Vasc Biol* 2003; 23: 695-701.
- [75] Szekanecz Z, Kerekes G, Soltész P. Vascular effects of biologic agents in RA and spondyloarthropathies. *Nat Rev Rheumatol* 2009; 5: 677-84.
- [76] Goebel KM, Storck U, Neurath. Intrasynovialorgote in therapy in rheumatoid arthritis. *Lancet* 1981; 1: 1015-17.
- [77] Arii K, Kumon Y, Ikeda Y, Suehiro T, Hashimoto K. Edaravone inhibits the disease activity in rheumatoid arthritis. *J Clin Pharm Ther* 2006; 31: 197-9.
- [78] Salvemini D, Cuzzocrea S. Therapeutic potential of superoxide dismutase mimetics as therapeutic agents in critical care medicine. *Crit Care Med* 2003; 31: S29-38.
- [79] Afonso V, Champy R, Mitrovic D, Collin P, Lomri A. Reactive oxygen species and superoxide dismutases: role in joint diseases. *Joint Bone Spine* 2007; 74: 324-9.
- [80] Ohno K, Ito M, Ichihara M. Molecular hydrogen as an emerging therapeutic medical gas for neurodegenerative and other diseases. *Oxid Med Cell Longev* 2012; doi: 10.1155/2012/353152
- [81] Nakao A, Toyoda Y, Sharma P, Evans M, Guthrie N. Effectiveness of hydrogen rich water on antioxidant status of subjects with potential metabolic syndrome--an open label pilot study. *J Clin Biochem Nutr* 2010; 46: 140-9.
- [82] Nakayama M, Kabayama S, Nakano H, *et al.* S. Biological effects of electrolyzed water in hemodialysis. *Nephron Clin Pract* 2009; 112: c9-15.
- [83] Kajiyama S, Hasegawa G, Asano M, *et al.* Supplementation of hydrogen-rich water improves lipid and glucose metabolism in patients with type 2 diabetes or impaired glucose tolerance. *Nutr Res* 2008; 28: 137-43.
- [84] Ito M, Ibi T, Sahashi K, Ichihara M, Ohno K. Open-label trial and randomized, double-blind, placebo-controlled, crossover trial of hydrogen-enriched water for mitochondrial and inflammatory myopathies. *Med Gas Res* 2011; 1: 24.
- [85] Ono H, Nishijima Y, Adachi N, *et al.* Improved brain MRI indices in the acute brain stem infarct sites treated with hydroxyl radical scavengers, Edaravone and hydrogen, as compared to Edaravone alone. A non-controlled study. *Med Gas Res* 2011; 1: 12.
- [86] Abraini JH, Gardette-Chauffour MC, Martinez E, Rostain JC, Lemaire C. Psychophysiological reactions in humans during an open sea dive to 500 m with a hydrogen-helium-oxygen mixture. *J Appl Physiol* 1994; 76: 1113-8.
- [87] Fontanari P, Badier M, Guillot C, *et al.* Changes in maximal performance of inspiratory and skeletal muscles during and after the 7.1-MPa Hydra 10 record human dive. *Eur J Appl Physiol* 2000; 81: 325-8.
- [88] Nakai Y, Sato B, Ushima S, Okada S, Abe K, Arai S. Hepatic oxidoreduction-related genes are upregulated by administration of hydrogen-saturated drinking water. *Biosci Biotechnol Biochem* 2011; 75: 774-6.